

### STOCK SOLUTION

10g NaNO<sub>3</sub> in 400 ml RO H<sub>2</sub>O  
1g CaCl<sub>2</sub> in 400 ml RO H<sub>2</sub>O  
3g K<sub>2</sub>HPO<sub>4</sub> in 400 ml RO H<sub>2</sub>O  
7g KH<sub>2</sub>PO<sub>4</sub> in 400 ml RO H<sub>2</sub>O  
3g MgSO<sub>4</sub>\*7H<sub>2</sub>O in 400 ml RO H<sub>2</sub>O  
1g NaCl in 400 ml RO H<sub>2</sub>O

Store prepped stock solutions in media storage bottles.

### NITSCH'S MICRONUTRIENTS

To 1L of RO H<sub>2</sub>O, add the following:

0.5 mL H<sub>2</sub>SO<sub>4</sub> (conc.)  
3 g MnSO<sub>4</sub> \* 4 H<sub>2</sub>O **or** 2.27g MnSO<sub>4</sub> \* 1 H<sub>2</sub>O  
0.5 g ZnSO<sub>4</sub> \* 7 H<sub>2</sub>O  
0.5 g H<sub>3</sub>BO<sub>4</sub>  
25 mg CuSO<sub>4</sub> \* 5H<sub>2</sub>O  
25 mg CoCl<sub>2</sub>  
25 mg Na<sub>2</sub>MoO<sub>4</sub> \* 2H<sub>2</sub>O

Store prepped stock solution in media storage bottle.

### ADDITIONAL COMPONENTS

0.7% agar  
1% FeCl<sub>3</sub> solution

### MEDIA PREPARATION

For 1L of solution:

1. Add 10mL of each stock solution to 1L volumetric flask.
2. Add 2mL of micronutrient solution to flask.
3. Add 1 drop of 1% FeCl<sub>3</sub> solution to flask (Do not add if non-culture grade reagents are used).
4. Add 7g agar to flask.
5. Bring flask to volume with RO H<sub>2</sub>O. Add H<sub>2</sub>O gradually and swirl flask to suspend agar in solution.
6. Transfer solution to 1L media storage bottle, swirling flask occasionally to resuspend agar. Cap bottle loosely (see next step).
7. Autoclave bottle and solution for ~30 min. If cap is on tight, the bottle will break in autoclave and spill solution. Keep bottle in autoclave for ~45min following cycle to allow bottle to cool to 50-60°C.
8. While media is in autoclave, lay out 25-30 (number needed varies based on amount used) 100 x 25mm Petri dishes along edge of lab desk. Place lids next to plates.
9. Remove media from autoclave and pour into Petri dishes. Fill each dish ~2/3 of the way. Replace lids on dishes – they should steam up. This will sterilize them. Allow dishes to cool on lab bench overnight. Once media has cooled and solidified, remove lids individually, remove any water that has condensed on the lid, and return lid. Excess water will interfere with spore sowing and germination. If dishes are not used immediately, place in crisper box and refrigerate.